# Personal details

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Name** | |  | | | |
| **Date** | |  | | | |
| **Email** | |  | | | |
| **Phone** | |  | | | |
| **Group** | |  | | | |
| Have you attended the Introductory Lecture? | | | | | | | |
| YES | |  | | NO |  | | |
| If *no*, when do you plan to attend? | | | | | |  | |

***For internal use***

|  |  |
| --- | --- |
| Meeting |  |
| Session I |  |
| Session II |  |
| Microscope |  |
| Email |  |
| All user list |  |
| Laser safety |  |
| Confirmation |  |
| Access |  |

# Experimental details

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| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Cell model** (yeast, bacteria, animal cells, cell line...) | | | | | | | | |
|  | | | | | | | | |
| **Sample description** (cell layer, tissue, embryo, whole animal...) | | | | | | | | |
|  | | | | | | | | |
| **Fixed samples** |  | | | | | | | |
| **Live imaging** |  | **CO2** | | | **Temperature** | |  | °C |
| **Imaging speed** (frames per s/min/h) | | | |  | | | | |
| **Time per experiment** (min, h, over night...) | | | | | |  | | |
| **Dyes and their excitation/emission maxima** (e.g., GFP 488/523) | | | | | | | | |
|  | | | | | | | | |
| **Multi-color** | if *yes*, how many simultaneously: | | | | | | | |
| **Photomanipulation** | if *yes*, at which wavelength: | | | | | | | |
| **Sample format** | Cover glass | | Plastic bottom | | | |  | |
|  | Slides | | Chambered coverglass | | | | Multiwell | |
|  | 35-mm dishes | | Chamber slides | | | |  | |
|  | Other: | |  | | | |  | |

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| **Resolution requirements** (leave blank if you don't know) | |
| Widefield (250-500 nm xy, 600-800 nm b z) |  |
| Confocal (160-300 nm xy, 500 nm z) |  |
| Super-resolution (< 160 nm xy, 500 nm z) |  |
| ***Note: super-resolution requires special dyes. Preferred:*** | |
| Abberior STAR RED/ORANGE, STAR 580/600/635/635P, Alexa Fluor 594/647, ATTO 590/633/647N | |

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| **Briefly describe your experimental strategy** (add comments or requests as needed) |
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